

Dichlorobis(sarcosinato)zinc(II)

M. Subha Nandhini,^a R. V. Krishnakumar^b and S. Natarajan^{a*}

^aDepartment of Physics, Madurai Kamaraj University, Madurai 625 021, India, and

^bDepartment of Physics, Thiagarajar College, Madurai 625 009, India

Correspondence e-mail: xray@mrna.tn.nic.in

Key indicators

Single-crystal X-ray study

$T = 293\text{ K}$

Mean $\sigma(\text{C}-\text{C}) = 0.004\text{ \AA}$

R factor = 0.028

wR factor = 0.075

Data-to-parameter ratio = 15.9

For details of how these key indicators were automatically derived from the article, see <http://journals.iucr.org/e>.

In the title compound, $[\text{ZnCl}_2(\text{CH}_3\text{NH}_2^+\text{CH}_2\text{COO}^-)_2]$, the sarcosine ligand coordinates to the metal in the zwitterionic form. The Zn atom has a distorted tetrahedral coordination with two Cl atoms and two O atoms, one each from the two crystallographically independent sarcosine molecules in the asymmetric unit. Head-to-tail hydrogen bonds between the amino acid molecules, in addition to $\text{N}-\text{H}\cdots\text{Cl}$, $\text{C}-\text{H}\cdots\text{O}$ and $\text{C}-\text{H}\cdots\text{Cl}$ hydrogen bonds, stabilize the packing of the molecules.

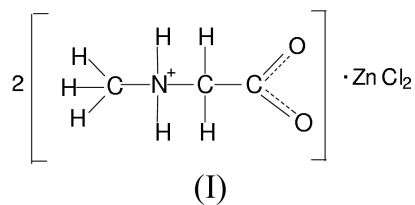
Received 29 August 2001

Accepted 10 September 2001

Online 20 September 2001

Comment

Sarcosine (*N*-methylglycine, $\text{CH}_3\text{NH}_2^+\text{CH}_2\text{COO}^-$), an intermediate in the metabolism of choline, occurs naturally in large amounts in starfish and sea urchins. The crystal structure of sarcosine itself was previously determined in our laboratory (Mostad & Natarajan, 1989). Zinc–amino acid complexes are interesting as zinc is known to compete successfully with cadmium for protein binding sites. Zinc also plays an important biological role in the formation of a structural motif called zinc fingers which are characteristic of certain proteins that bind to DNA. Recently, the crystal structure of a complex of sarcosine with zinc chloride, namely trichloro(sarcosinio)-zinc(II) monohydrate (Krishnakumar *et al.*, 2001), in which the amino acid exhibits an unusual cationic form, was reported. The present work reports the crystal structure of a complex, (I), of sarcosine with zinc chloride where the amino acid adopts the usually expected zwitterionic form. The crystal structures of complexes of ZnCl_2 with glycine (Hariharan *et al.*, 1989), *L*-proline (Yukawa *et al.*, 1985) and *L*-histidine (Foster *et al.*, 1993) have already been reported.



In Fig. 1, the molecular structure and atom-numbering scheme are shown. Both the sarcosine molecules in the asymmetric unit exist as zwitterions, each with a positively charged amino group and a negatively charged carboxylate group. However, the $\text{C1}-\text{O1}$ and $\text{C1}-\text{O2}$ bond lengths in molecule *A* [1.276 (3) and 1.223 (3) Å, respectively] and molecule *B* [1.261 (3) and 1.213 (3) Å, respectively] show significant deviations from those usually exhibited by a zwitterion. The larger value observed in the $\text{C1}-\text{O1}$ bond lengths may be due to the fact that both the O1 atoms of molecule *A*

and *B* participate in the coordination environment around the metal. The angle between the planes formed by the atoms of the two sarcosine molecules coordinating to Zn is 81.5 (1)°.

Zinc is known to have both tetrahedral and octahedral coordination in crystal structures (Cingi *et al.*, 1972). In the present structure, zinc has a distorted tetrahedral environment with two Cl atoms and two carboxyl O atoms, one each from the two crystallographically independent sarcosine molecules in the asymmetric unit. The angles around the Zn atom range from 101.36 (5) to 115.74 (3)°. The coordination environment around Zn in trichloro(sarcosinio)zinc(II) monohydrate, however, is different since it involves three chlorines and one of the carboxyl O atoms.

Head-to-tail hydrogen bonds between the amino acid molecules, in addition to N—H···Cl, C—H···O and C—H···Cl hydrogen bonds, stabilize the three-dimensional network of the molecules. The crystal structure of (I) does not bear any relation either to that of trichloro(sarcosinio)zinc(II) monohydrate or to those of sarcosine cadmium chloride tetrahydrate (Krishnakumar *et al.*, 1996) and sarcosine barium chloride tetrahydrate (Krishnakumar & Natarajan, 1995). However, the unit-cell parameters of (I) and those of sarcosine barium chloride tetrahydrate bear an interesting relationship as the *b* and *c* axes are very nearly equal and the cell length *a* in (I) is twice that of the other.

Experimental

Colorless single crystals of (I) were grown as transparent needles from a saturated water–acetone mixture containing sarcosine and zinc chloride in a 1:1 stoichiometric ratio.

Crystal data

[ZnCl₂(C₆H₁₄N₂O₄)]
M_r = 314.46
 Orthorhombic, *Pbca*
a = 14.191 (2) Å
b = 10.655 (1) Å
c = 15.917 (2) Å
V = 2406.5 (4) Å³
Z = 8
D_x = 1.736 Mg m⁻³
D_m = 1.74 Mg m⁻³

D_m measured by flotation in a mixture of carbon tetrachloride and bromoform
 Cu Kα radiation
 Cell parameters from 25 reflections
 $\theta = 15\text{--}27^\circ$
 $\mu = 6.94\text{ mm}^{-1}$
T = 293 (2) K
 Needle, colorless
 0.16 × 0.12 × 0.10 mm

Data collection

Enraf–Nonius CAD-4 diffractometer
 ω – 2θ scans
 Absorption correction: ψ scan (North *et al.*, 1968)
T_{min} = 0.400, *T_{max}* = 0.503
 2182 measured reflections
 2182 independent reflections

2004 reflections with *I* > 2σ(*I*)
 $\theta_{\text{max}} = 67.9^\circ$
h = 0 → 16
k = 0 → 12
l = 0 → 19
 2 standard reflections every 200 reflections
 intensity decay: 0.1%

Refinement

Refinement on *F*²
R[*F*² > 2σ(*F*²)] = 0.028
wR(*F*²) = 0.075
S = 1.18
 2182 reflections
 137 parameters
 H-atom parameters constrained

$w = 1/[\sigma^2(F_o^2) + (0.0299P)^2 + 2.1069P]$
 where $P = (F_o^2 + 2F_c^2)/3$
 $(\Delta/\sigma)_{\text{max}} = 0.001$
 $\Delta\rho_{\text{max}} = 0.28\text{ e \AA}^{-3}$
 $\Delta\rho_{\text{min}} = -0.43\text{ e \AA}^{-3}$
 Extinction correction: *SHELXL97*
 Extinction coefficient: 0.00071 (4)

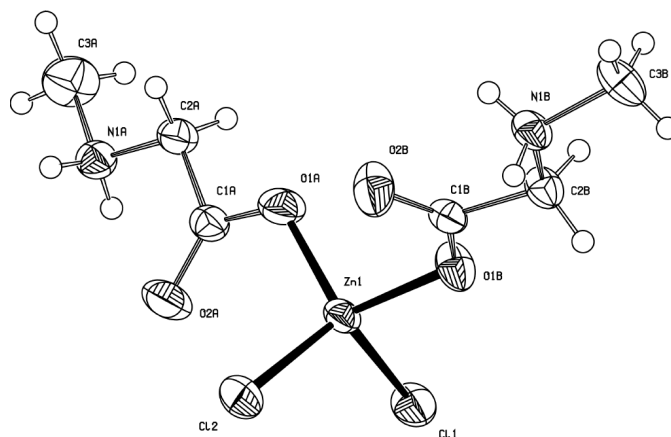


Figure 1
 The structure of (I) with 50% probability displacement ellipsoids.

Table 1

Selected geometric parameters (Å, °).

Zn1—O1A	1.9484 (19)	O2B—C1B	1.213 (3)
Zn1—O1B	1.9713 (17)	N1A—C2A	1.471 (3)
Zn1—Cl2	2.2595 (7)	N1A—C3A	1.484 (3)
Zn1—Cl1	2.2739 (7)	N1B—C3B	1.478 (3)
O1A—C1A	1.276 (3)	N1B—C2B	1.479 (3)
O1B—C1B	1.261 (3)	C1A—C2A	1.514 (3)
O2A—C1A	1.223 (3)	C1B—C2B	1.519 (3)
O1A—Zn1—O1B	107.20 (8)	O2A—C1A—O1A	126.6 (2)
O1A—Zn1—Cl2	113.40 (6)	O2A—C1A—C2A	120.0 (2)
O1B—Zn1—Cl2	111.71 (6)	O1A—C1A—C2A	113.4 (2)
O1A—Zn1—Cl1	106.43 (6)	O2B—C1B—O1B	127.0 (2)
O1B—Zn1—Cl1	101.36 (5)	O2B—C1B—C2B	119.3 (2)
Cl2—Zn1—Cl1	115.74 (3)	O1B—C1B—C2B	113.7 (2)
C2A—N1A—C3A	113.9 (2)	N1A—C2A—C1A	111.4 (2)
C3B—N1B—C2B	112.97 (19)	N1B—C2B—C1B	111.88 (19)
C3A—N1A—C2A—C1A	−171.6 (2)	C3B—N1B—C2B—C1B	−166.3 (2)
O2A—C1A—C2A—N1A	−1.3 (3)	O2B—C1B—C2B—N1B	2.1 (3)
O1A—C1A—C2A—N1A	178.0 (2)	O1B—C1B—C2B—N1B	−179.2 (2)

Table 2

Hydrogen-bonding geometry (Å, °).

<i>D</i> —H··· <i>A</i>	<i>D</i> —H	H··· <i>A</i>	<i>D</i> ··· <i>A</i>	<i>D</i> —H··· <i>A</i>
N1A—H1A1···O2B ⁱ	0.90	1.93	2.714 (3)	144
N1A—H1A2···Cl1 ⁱⁱ	0.90	2.35	3.223 (2)	163
N1B—H1B1···Cl2 ⁱⁱⁱ	0.90	2.42	3.291 (2)	164
N1B—H1B2···Cl2 ^{iv}	0.90	2.44	3.191 (2)	141
N1B—H1B2···O2A ⁱⁱⁱ	0.90	2.48	2.985 (3)	116
C2A—H2A2···O1B ^v	0.97	2.40	3.342 (3)	164
C2B—H2B2···Cl1 ^{vi}	0.97	2.76	3.610 (3)	146
C3A—H3A1···O2A ⁱⁱ	0.96	2.66	3.555 (4)	156

Symmetry codes: (i) 2 − *x*, 1 − *y*, 1 − *z*; (ii) $\frac{3}{2} - x, \frac{1}{2} + y, z$; (iii) $\frac{1}{2} + x, \frac{1}{2} - y, 1 - z$; (iv) 2 − *x*, −*y*, 1 − *z*; (v) 2 − *x*, $\frac{1}{2} + y, \frac{1}{2} - z$; (vi) $\frac{1}{2} + x, y, \frac{1}{2} - z$.

The H atoms were placed in calculated positions and were allowed to ride on their respective parent atoms using *SHELXL97* (Sheldrick, 1997) defaults.

Data collection: *CAD-4 Software* (Enraf–Nonius, 1989); cell refinement: *CAD-4 Software*; data reduction: *CAD-4 Software*;

program(s) used to solve structure: *SHELXS97* (Sheldrick, 1990); program(s) used to refine structure: *SHELXL97* (Sheldrick, 1997); molecular graphics: *PLATON* (Spek, 1999); software used to prepare material for publication: *SHELXL97*.

The authors thank the Council of Scientific and Industrial Research (CSIR), India, for financial assistance. The authors also thank the UGC for the DRS programme and the Bioinformatics Centre, Madurai Kamaraj University, for providing the Cambridge Structural Database (Allen & Kennard, 1993).

References

- Allen, F. H. & Kennard, O. (1993). *Chem. Des. Autom. News*, **8**, 1, 31–37.
- Cingi, M. B., Guastini, C., Musatti, A. & Nardelli, M. (1972). *Acta Cryst.* **B28**, 667–672.
- Enraf–Nonius (1989). *CAD-4 Software*. Version 5.0. Enraf–Nonius, Delft, The Netherlands.
- Foster, M., Burth, R., Powell, A. K., Eiche, P. & Vahrenkamp, H. (1993). *Chem. Ber.* **126**, 2643–2648.
- Hariharan, M., Rajan, S. S., Srinivasan, R. & Natarajan, S. (1989). *Z. Kristallogr.* **188**, 217–222.
- Krishnakumar, R. V., Subha Nandhini, M. & Natarajan, S. (2001). *Acta Cryst.* **E57**, m192–m194.
- Krishnakumar, R. V. & Natarajan, S. (1995). *Cryst. Res. Technol.* **30**, 825–830.
- Krishnakumar, R. V., Rameela, M. P. & Natarajan, S. (1996). *Cryst. Res. Technol.* **31**, 203–207.
- Mostad, A. & Natarajan, S. (1989). *Acta Chem. Scand.* **43**, 1004–1006.
- North, A. C. T., Phillips, D. C. & Mathews, F. S. (1968). *Acta Cryst.* **A24**, 351–359.
- Sheldrick, G. M. (1990). *Acta Cryst.* **A46**, 467–473.
- Sheldrick, G. M. (1997). *SHELXL97*. University of Göttingen, Germany.
- Spek, A. L. (1999). *PLATON for Windows*. Utrecht University, The Netherlands.
- Yukawa, Y., Yasukawa, N., Inomata, Y. & Takeuchi, T. (1985). *Bull. Chem. Soc. Jpn.* **58**, 1591–1592.